

# A forage-only diet alters the metabolic response of horses in training

A. Jansson<sup>†</sup> and J. E. Lindberg

Department of Animal Nutrition and Management, Swedish University of Agricultural Sciences, SE-750 07 Uppsala, Sweden

(Received 2 September 2011; Accepted 26 March 2012; First published online 4 May 2012)

*Most athletic horses are fed a high-starch diet despite the risk of health problems. Replacing starch concentrate with high-energy forage would alleviate these health problems, but could result in a shift in major substrates for muscle energy supply from glucose to short-chain fatty acids (SCFA) due to more hindgut fermentation of fibre. Dietary fat inclusion has previously been shown to promote aerobic energy supply during exercise, but the contribution of SCFA to exercise metabolism has received little attention. This study compared metabolic response with exercise and lactate threshold ( $V_{La4}$ ) in horses fed a forage-only diet (F) and a more traditional high-starch, low-energy forage diet (forage–concentrate diet - FC). The hypothesis was that diet F would increase plasma acetate concentration and increase  $V_{La4}$  compared with diet FC. Six Standardbred geldings in race training were used in a 29-day change-over experiment. Plasma acetate, non-esterified fatty acids (NEFA), lactate, glucose and insulin concentrations and venous pH were measured in samples collected before, during and after a treadmill exercise test (ET, day 25) and muscle glycogen concentrations before and after ET. Plasma acetate concentration was higher before and after exercise in horses on diet F compared with diet FC, and there was a tendency ( $P = 0.09$ ) for increased  $V_{La4}$  on diet F. Venous pH and plasma glucose concentrations during exercise were higher in horses on diet F than diet FC, as was plasma NEFA on the day after ET. Plasma insulin and muscle glycogen concentrations were lower for diet F, but glycogen utilisation was similar for the two diets. The results show that a high-energy, forage-only diet alters the metabolic response to exercise and, with the exception of lowered glycogen stores, appears to have positive rather than negative effects on performance traits.*

**Keywords:** acetate, blood pH, exercise, insulin, muscle glycogen

## Implication

It is a serious animal welfare issue that diets associated with gastrointestinal disorders and abnormal and stereotypical behaviour are fed to horses today. There is an urgent need for diets that support the natural digestive function and behaviour of horses. Such diets would reduce welfare problems and healthcare costs for the horse industry.

## Introduction

Current practice in feeding performance horses worldwide is to use fairly late-cut forage, whereas, for example, in dairy production early-cut, high-energy forage is used. The high-energy requirements of many performance horses means that supplementation with more energy-dense feeds is necessary and more than 40% of the diet commonly consists of cereal-based, high-starch concentrates (Glade, 1983;

Redbo *et al.*, 1998; Williamson *et al.*, 2007). This is a serious animal welfare issue, as low forage to concentrate ratios and high starch intake are associated with reduced gut microbial stability (Willing *et al.*, 2009), gastrointestinal disorders (Tinker *et al.*, 1997; Hudson *et al.*, 2001; Luthersson *et al.*, 2009) and abnormal and stereotypical behaviour (Gillham *et al.*, 1994; Redbo *et al.*, 1998; Waters *et al.*, 2002). In addition, rhabdomyolysis has been associated with high concentrate intakes (MacLeay *et al.*, 1999). We have shown in earlier studies that Standardbred horses in training can maintain BW and condition on high-energy, forage-only diets (Connysson *et al.*, 2006 and 2010; Muhonen *et al.*, 2009). We found no detrimental effect of increased CP intake (which may be associated with the use of high-energy forages) on plasma lactate concentration and pH during exercise, but urinary pH decreased and evaporative losses tended to increase (Connysson *et al.*, 2006). However, the effect on exercise performance of forage-only diets compared with the traditional forage–concentrate diet (FC) has not yet been studied.

<sup>†</sup> E-mail: Anna.Jansson@slu.se

From a physiological point of view, the horse is adapted to continuous grazing of a forage-only diet (low in starch content) and has a well-developed symbiosis with the hindgut microbiota fermenting the forage fibres. This results in the production of energy-yielding substrates in the form of short-chain fatty acids (SCFA), and the proportion of acetate increases and propionate decreases when forage : concentrate ratio is increased (Hintz *et al.*, 1971; Willard *et al.*, 1977). This suggests that horses are well adapted to rely on fat metabolism and aerobic energy supply at rest, but probably also during exercise. The adaptation to, and importance of, aerobic substrate utilisation during exercise in Thoroughbred and Standardbred horses has been documented in a number of studies (Lindholm and Piehl, 1974; Wilson *et al.*, 1987; Essén-Gustavsson *et al.*, 1989) and is also confirmed by the correlation between the performance of Standardbred horses and plasma lactate threshold ( $V_{La4}$ ) (Persson, 1983). Equine muscle also shows high plasticity with respect to its adaptation to energetic demands as a result of exercise training (Voiton *et al.*, 2007). Therefore, it appears reasonable to assume that the substrate profile for efficient muscle energy metabolism in horses will comprise glucose, long-chain fatty acid (LCFA) from the diet or from body lipid stores (Pagan *et al.*, 2002; Geor, 2006) and SCFA from hindgut fermentation of dietary fibre (Palmgren-Karlsson *et al.*, 2002). The aim of this study was to compare the effects of a high-energy forage-only diet with those of a 50:50 (dry matter (DM) basis) forage : concentrate (starch-rich) diet in terms of lactate threshold ( $V_{La4}$ ), muscle glycogen concentration and metabolic plasma profile in Standardbred horses in training. The hypothesis was that the forage-only diet would increase plasma acetate concentration and  $V_{La4}$ .

## Material and methods

### Horses

Six Standardbred geldings in race training were used, which were aged  $6.5 \pm 0.4$  years (mean  $\pm$  s.d.). The average number of races in which the horses had competed was  $27 \pm 8$ , and the average racing record was  $77.3 \pm 0.8$  s/1000 m. They had an initial BW of  $515 \pm 21$  kg. The horses were kept at a training camp for harness racing 20 km south of Uppsala, Sweden. They were housed in individual stalls on wood shavings during the night and were kept together in a sand/clay paddock between 0800 and 1500 h on days without training. All horses had passed a flexure test before the study and were regarded as healthy. The experiment was approved by the Uppsala local ethics committee and was conducted in the period October–December 2007.

### Experimental design

**Diets.** The horses were offered a forage-only diet (F) consisting of early-cut haylage (timothy, meadow fescue mixture; Table 1) and a mixed diet (FC) consisting of late-cut haylage (timothy, meadow fescue mixture) supplemented with concentrate (50:50 DM basis) in a 29-day change-over design experiment. Feed allowance was based on individual BW and

**Table 1** DM (%), estimated energy (MJ ME/kg DM), chemical (g/kg DM) and microbial composition (cfu/g fresh matter) of feeds in the experimental diets

	Forage		Concentrate
	Diet F	Diet FC	Diet FC
DM	80	78	90
ME	10.4	8.8	11.4
Ash	75	56	37
CP	104	61	174
NDF	605	600	203
ADF	363	370	117
Lignin	51	69	27
Crude fat	19	15	60
WSC	79	147	
Free glucose	35	22	3
Free fructose	31	57	0
Fructans	4	51	5
Starch	0	0	358
Yeast	$\leq 2.5$	$\leq 3.6$	–
Mould	$< 2.0$	$\leq 2$	–

DM = dry matter; ME = metabolisable energy; cfu = colony-forming units; F = forage-only diet; FC = forage–concentrate diet; WSC = water-soluble carbohydrates.

was 13 to 17.4 kg haylage and 180 to 240 g sugar (only to ensure complete intake of the salt, mineral and vitamin supplements) for diet F and 6.3 to 8.4 kg haylage, 5.3 to 7.1 kg oats, 0.9 to 1.2 kg soy bean meal, 0.18 to 0.24 kg wheat bran and 90 to 120 g sugar for diet FC. The diets were estimated to be iso-caloric and iso-nitrogenous, and provided energy and nutrients according to requirements specified by National Research Council (NRC, 1989). Horses on both diets were offered a mineral and vitamin supplement ( $51 \pm 2$  g/day, Miner Röd, Krafft, Falkenberg, Sweden), NaCl ( $36 \pm 1$  g/day), and those on diet FC were offered ground chalk (calcium carbonate,  $34 \pm 1$  g/day) in order to meet mineral and vitamin requirements specified by NRC (1989). Water was provided *ad libitum* from graded buckets. The forage allowance was fed in the afternoon and the concentrate and mineral and vitamin supplement (diet FC) at 1500, 2300 and 0600 h. With diet F, the mineral and vitamin supplement was fed at 2300 and 0600 h. Diet FC was introduced gradually during the experimental period (on days 1 and 2, horses were fed 50% of the F diet and 50% of the FC diet, and then the FC diet was increased by 10% per day until the full ration was reached on day 7). Diet F was introduced abruptly on day 1.

**Training.** The horses were given sub-maximal warm-up (3.5 to 5 km, heart rate  $< 200$  beats/min) and intensive training (heart rate  $> 200$  beats/min) on an oval or straight field track ( $\sim 0.6\%$  incline) on days 2, 5, 7, 9, 13, 17, 21, 25 and 29 in order to maintain but not improve fitness. The intensive training consisted of interval training (four intervals of 600 m) or 1600 to 2000 m heats, and the training protocol was exactly the same in both periods.

**Exercise test (ET).** On day 25 of each experimental period, the horses performed a standardised incremental ET on a treadmill (Säto, Knivsta, Sweden) located at a veterinary clinic 25 km from the training camp. The horses were transported to the clinic by trailer. All horses had prior experience of exercise on this treadmill (minimum two occasions).

Before ET, the horses were kept together in the paddock from 0730 to 1030 h and were then offered 1 kg of forage (diet F) and 1 kg of oats (diet FC). After this, water but no feed was offered until the horses returned to the stable at about 2400 h. This design was chosen because practical experience indicated that many horses do not consume any feed while at the race-track. On the day after ET, all horses were fed at 0600 h as usual, but were kept in their stalls until 1100 h.

Exercise testing consisted of two phases. The first phase (warm-up, walk at 2 m/s for 3 min, trot 6 m/s for 5 min, trot 9.5 m/s for 5 min and walk 2 m/s for 3 min, no incline) was performed 25 min before the second phase, which consisted of 5 min of walk (0% incline) and then a stepwise increase in velocity (6.0, 7.0, 8.0, 9.0, 9.5 m/s) at an incline of 6.3% every second minute until the horses reached or exceeded a heart rate of 200 beats/min. Four horses reached 9.0 m/s and two reached 9.5 m/s. The horses walked for 5 min on the treadmill after the final step.

#### Sampling

**Feed and water intake, BW and condition.** Samples of concentrate were collected from each batch used in every period and samples of forage from each new bale opened and kept frozen at  $-20^{\circ}\text{C}$  until analysis. The forage samples were pooled and analysed as one sample per feed and period. Feed leftovers were weighed and eliminated every day. Daily nutrient and energy intake was calculated using feed intake data and analysis of feeds and leftovers. Water intake (drinking) was measured on days 19 to 28.

BW was recorded before each training session, before the warm-up prior to ET, immediately after ET, on the day after ET (at 1100 h) and on days 27 to 29 (afternoon). Changes in body condition were assessed by a simple recording of whether the ribs were easy to palpate or not and whether they were visible during motion.

**ET.** Heart rate was recorded by a heart frequency meter (Polar RS800, Kempele, Finland) during exercise and manually post exercise with a stethoscope. Heart rate was measured before warm-up, during warm-up (9.5 m/s), immediately after warm-up, after 25 min of rest, during each incremental step, after 5 min walking on the treadmill and 10, 25, 55 and 85 min after the end of walking.

Breathing frequency was recorded before warm-up, directly after warm-up, after 25 min of rest, after 5 min walking on the treadmill and 10, 25, 55 and 85 min after the end of walking. Rectal temperature was recorded before warm-up, directly after warm-up, after 25 min of rest and 10, 25, 55, 85 min after the end of walking.

A jugular catheter was introduced under local anaesthesia (Carbocain 20 mg/ml, AstraZeneca AB, Sweden) before

transport to the clinic, and blood samples (20 ml/sample) were collected in Li-heparinised tubes before warm-up, immediately after warm-up, after 25 min of rest, at the end of each incremental step, after 5 min walking on the treadmill and 10, 25, 55 and 85 min after the end of walking. A final blood sample was taken on the day after ET, at 1100 h. The blood samples were kept chilled until centrifugation and frozen at  $-20^{\circ}\text{C}$  for later analysis. Samples collected for analysis of blood pH, and  $\text{TCO}_2$  and  $\text{HCO}_3$  were analysed within 10 min of collection.

Muscle biopsies were taken from *m. gluteus medius* at a depth of  $\sim 6$  cm according to the method described by Lindholm and Piehl (1974). A local anaesthetic (Carbocain 20 mg/ml, AstraZeneca) was applied to the area and a nose twitch was used. Biopsies were taken before warm-up and immediately after ET, frozen in liquid nitrogen and stored at  $-80^{\circ}\text{C}$  until analysis.

#### Analyses

Preparation and conventional chemical analyses (DM, nitrogen, NDF, ADF, lignin, water-soluble carbohydrate (WSC) and ash) of feeds were performed as described by Palmgren-Karlsson *et al.* (2000). Minerals were analysed with inductively coupled plasma optical emission spectrometry (ICP-OES; SS-EN 14538:2006, Ametek Spectro, Kleve, Germany). The metabolisable energy content of the forage was estimated by an *in vitro* method (Lindgren, 1979).

**Analyses of plasma, blood and muscle.** Plasma insulin was analysed using an ELISA method (Mercodia equine insulin kit, Mercodia, Uppsala, Sweden). For quantitative determination of non-esterified fatty acids (NEFA), an enzymatic colorimetric method was used (ACS-ACOD method, Wako Chemicals GmbH, Neuss, Germany). Plasma urea, acetate and glucose concentrations were analysed with an enzymatic colorimetric/UV method (Boehringer Mannheim/R-Biopharm, Darmstadt, Germany), and lactate was analysed using an ELISA method (R-Biopharm GmbH, Darmstadt, Germany). Venous pH,  $\text{TCO}_2$  and  $\text{HCO}_3$  were analysed using an i-STAT<sup>®</sup> 1 analyser (Abbott Laboratories, Abbott Park, Illinois, USA). Total plasma protein (TPP) concentration was measured using a refractometer (Atago, Sur-Ne, Tokyo, Japan) in samples taken before warm-up, before ET, 55 min after walking and the day after walking.

Muscle samples for glycogen analysis were freeze-dried and dissected free from visible blood, connective tissue and fat under a microscope before analysis. A sample of about 1 to 2 mg muscle fibre was boiled in 1 M HCl, and glucose residues were determined by fluorescence (Lowry and Passonneau, 1973).

#### Calculations and statistical analysis

The velocity at plasma lactate concentration 4 mmol/l ( $V_{\text{La4}}$ ) was calculated individually from an exponential curve fitted by Microsoft Office Excel 2007.

All data were subjected to analysis of variance (GLM procedure in the Statistical Analysis Systems package 9.1; SAS Institute Inc., Cary, NC, USA) using the following model:  $Y_{ijk} = \mu + \alpha_i + \beta_j + \gamma_k + \varepsilon_l + (\beta\gamma)_{jk} + e_{ijkl}$ , where  $Y_{ijk}$  is

the observation,  $\mu$  the mean value,  $\alpha_i$  the effect of animal,  $\beta_j$  the effect of diet,  $\gamma_k$  the effect of sample,  $\varepsilon_l$  the effect of period,  $(\beta\gamma)_{jk}$  the effect of interaction between diet and sample and  $e_{ijkl}$  the residuals;  $e_{ijkl} \sim \text{IND}(0, \delta^2)$ . The value used for significance within and between treatments was  $P < 0.05$ . Values are presented as least square mean  $\pm$  s.e.m.

**Results**

*General observations*

All horses completed the study. No health problems were observed, except in one horse that developed a fever 2 days into period 1 (resulting in three missed training sessions). There were leftovers on both diets (Table 2), and forage intake corresponded to 69% of the allowance on both diets, resulting in a forage : concentrate ratio of 40 : 60 for diet FC. A comparison of the *in vitro* digestible organic matter (IVDOM) of the individual leftovers and the feed samples collected on days 20 to 22 showed consistently lower IVDOM in the leftovers compared with the feed for diet F, but not diet FC (Diet F: feed = 78.3%, leftovers = 73.7% to 76.9%; diet FC: feed = 66.9%, leftovers = 63.7% to 69.6%), indicating selective feed intake in all individuals on diet F.

Body condition was similar in four individuals on both diets, but two individuals had higher body condition on diet FC (horse 1 changed from 'ribs visible at motion' to 'not visible but easy to palpate' and horse 2 changed from 'ribs not visible at motion but easy to palpate' to 'not visible, not easy to palpate'). Mean BW was higher ( $P = 0.0006$ ) for horses on diet F ( $519.0 \pm 0.5$  kg) than diet FC ( $516.0 \pm 0.5$  kg). Water intake was higher with diet F than diet FC ( $30.0 \pm 0.5$  v.  $19.0 \pm 0.5$  l/day,  $P < 0.0001$ ).

*ET*

There was no significant difference in  $V_{La4}$  between diets, although there was a tendency ( $P = 0.086$ ) for higher  $V_{La4}$  in

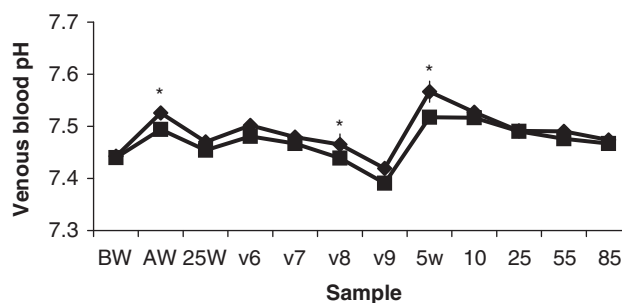
**Table 2** Daily feed allowance, feed intake (kg) and nutrient (g) and estimated metabolisable energy intake (MJ ME) during 29 days on a high-energy, forage-only diet (F) and a mixed forage-concentrate diet (FC) (LSmeans  $\pm$  s.e.)

	Diet F	Diet FC
Forage allowance	14.56 $\pm$ 0.04	7.00 $\pm$ 0.04
Forage intake	10.07 $\pm$ 0.09	4.88 $\pm$ 0.09
Concentrate allowance	0.29 $\pm$ 0.03	7.31 $\pm$ 0.03
Concentrate intake	0.29 $\pm$ 0.04	7.07 $\pm$ 0.04
CP intake <sup>1</sup>	1132 $\pm$ 87	1467 $\pm$ 66
NDF intake	6588 $\pm$ 507	3885 $\pm$ 270
Starch intake	0	2503 $\pm$ 108
WSC intake	861 $\pm$ 66	605 $\pm$ 60
Energy intake <sup>2</sup>	110 $\pm$ 6	116 $\pm$ 6

WSC = water-soluble carbohydrates.

<sup>1</sup>Corresponds to 113% to 146% of the requirements for very heavy exercise suggested by NRC (2007).

<sup>2</sup>Corresponds to 90% to 94% of the requirements for very heavy exercise suggested by NRC (2007).



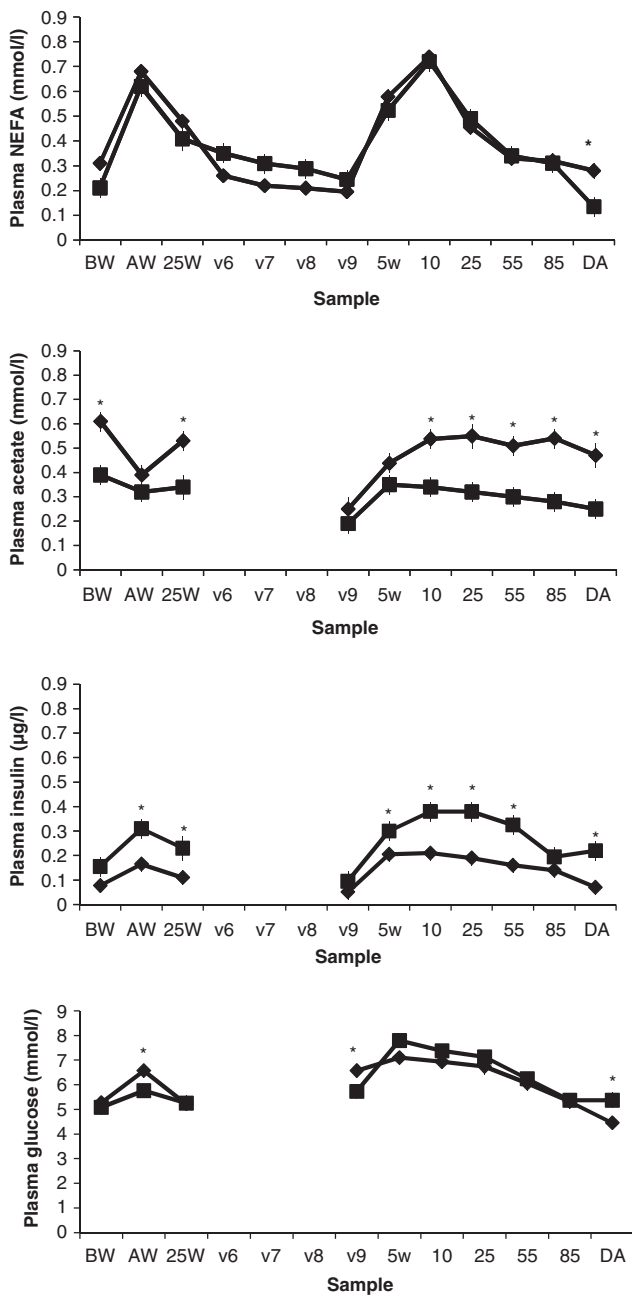
**Figure 1** Venous pH before, during and after an incremental exercise test. Values (LSmeans  $\pm$  SE) for six Standardbred geldings on a high-energy, forage-only diet (F; diamonds) and a 40:60 forage-concentrate diet (FC; squares). BW = before warm-up, AW = after warm-up, 25W = 25 min after warm-up, v6-v9 = incremental exercise test at velocities 6, 7, 8 and 9 m/s (treadmill incline 6.3%), 5w = after 5 min walk, 10, 25, 55, 85 min after the walk. The effect of diet was significant (ANOVA,  $P = 0.0001$ ). \*Indicates significant difference ( $P < 0.05$ ) between diets for single samples.

horses on diet F ( $8.0 \pm 0.1$  v.  $7.6 \pm 0.1$  m/s). Plasma lactate concentration was higher ( $P < 0.05$ ) in horses on diet FC compared with diet F at the end of the 5-min walk and 10 min after the walk on the treadmill (diet FC:  $8.1 \pm 0.4$  and  $4.4 \pm 0.4$  v. diet F:  $5.7 \pm 0.4$  and  $3.2 \pm 0.4$  mmol/l). Venous pH (Figure 1),  $\text{TCO}_2$  and  $\text{HCO}_3$  ( $P = 0.0001$ ) in connection with exercise were higher in horses on diet F compared with diet FC (pooled data for  $\text{TCO}_2$ :  $31.2 \pm 0.2$  v.  $30.3 \pm 0.2$  mmol/l and  $\text{HCO}_3$ :  $30.0 \pm 0.2$  v.  $29.1 \pm 0.2$  mmol/l). Acetate concentration was higher with diet F than diet FC ( $P = 0.0001$ ) and plasma NEFA concentration on the day after ET was higher ( $P < 0.05$ ) with diet F compared with diet FC (Figure 2). Plasma insulin concentration was higher ( $P = 0.0001$ ) in horses on diet FC compared with diet F (Figure 2), and glucose concentration was higher ( $P < 0.05$ ) with diet F immediately after warm-up and at 9 m/s, but lower ( $P < 0.05$ ) on the day after ET (Figure 2). Muscle glycogen content was higher ( $P < 0.05$ ) in horses on diet FC than diet F both before ( $644 \pm 22$  v.  $560 \pm 22$  mmol glucosyl units/kg dry weight) and after exercise ( $546 \pm 22$  v.  $473 \pm 22$  mmol glucosyl units/kg dry weight). There were no differences between the diets in heart rate ( $109 \pm 1$  and  $111 \pm 1$  beats/min in F and FC, respectively), breathing frequency ( $62 \pm 3$  and  $68 \pm 3$  beats/min in F and FC, respectively) or rectal temperature ( $38.5 \pm 0.1$  and  $38.5 \pm 0.1$  beats/min in F and FC, respectively), before, during and after exercise (pooled data).

There were no differences in BW between diets before (diet FC:  $509 \pm 1$  kg and diet F:  $511 \pm 2$  kg) and after warm-up and ET (diet FC:  $501 \pm 1.0$  kg and diet F:  $500 \pm 1$  kg). The pre-exercise BW was not recovered until day 28 on both diets. Mean TPP during ET day and on the day after was lower with diet F than diet FC ( $64.6 \pm 0.6$  v.  $67.0 \pm 0.6$  g/l).

**Discussion**

Horses fed the forage-only diet showed an altered metabolic response during exercise compared with horses on the forage-concentrate diet, and few, if any, effects likely to impair



**Figure 2** Plasma glucose<sup>a</sup>, insulin<sup>a</sup>, acetate<sup>a</sup> and non-esterified fatty acid (NEFA) concentrations before, during and after an incremental exercise test. Values (LSmeans  $\pm$  s.e.) from six Standardbred geldings on a high-energy, forage-only diet (F; diamonds) and a 40:60 forage-concentrate diet (FC; squares). BW = before warm-up, AW = after warm-up, 25W = 25 min after warm-up, v6–v9 = incremental exercise test at velocities 6, 7, 8 and 9 m/s (treadmill incline 6.3%), 5w = after 5 min walk, 10, 25, 55, 85 min after the walk and DA = day after at 1100 h. <sup>a</sup> = no analyses for v6–v8 available. The effect of diet was significant for acetate and insulin (ANOVA,  $P < 0.0001$ ). \*Indicates significant difference ( $P < 0.05$ ) between diets for single samples.

performance were observed. There was an increase in plasma acetate concentration on the F diet, as expected. Moreover, with this diet, plasma lactate concentration was significantly lower post exercise and there was a tendency for higher  $V_{La4r}$ , indicating that aerobic energy utilisation may

have improved. In addition, venous pH increased during exercise in horses on the F diet, which could counteract the acidosis induced by the exercise typically performed by racehorses. This indicates that high-energy, forage-only diets are an interesting alternative to conventional diets.

The increased plasma levels of acetate in horses on the F diet can be explained by more extensive hindgut fermentation due to higher fibre intake, and subsequent production of SCFA (Hintz *et al.*, 1971; Willard *et al.*, 1977). SCFA absorbed from the hindgut can be used as substrates in body tissue metabolism. Propionate may be primarily used to produce glucose in the gluconeogenic pathway, whereas acetate and butyrate can be transformed to acetyl-CoA and then utilised as substrates for aerobic energy metabolism in the tri-carboxylic pathway in the muscle (Voiton *et al.*, 2007). The latter also applies to LCFA that are metabolised through  $\beta$ -oxidation in the muscle. It has been suggested that long-term feeding of supplemental fat to exercising horses increases the mobilisation and speed of mobilisation of free fatty acids (FA), increasing the speed of uptake into muscle of free FA, lowering lactic acid production, imparting a glycogen-sparing effect and increasing pre-exercise muscle glycogen levels (Potter *et al.*, 1992; Harris and Harris, 2005). However, there is a large variation in the effects reported in the literature, and only a few studies report direct benefits on performance traits (Geor, 2006). Pagan *et al.* (2002) used a stable glucose isotope and showed that a high-fat diet can be glucose-sparing by increasing FA oxidation from body lipid stores during exercise, suggesting that performance could be improved. In the present study, no clear effect of the F diet on performance was observed, but there was a lowering of post-exercise plasma lactate concentration, confirming previous findings for a high-fibre diet (Palmgren-Karlsson *et al.*, 2002).

Waller and Lindinger (2007) reported profound plasma alkalosis, that is, a decrease in venous plasma hydrogen concentration and an increase in plasma  $\text{TCO}_2$  and  $\text{HCO}_3^-$ , after post-exercise oral administration of sodium acetate. These results were confirmed in the present study, where venous pH,  $\text{TCO}_2$  and  $\text{HCO}_3^-$  concentrations were higher in horses on the F diet. The alkalising effect of a forage-only diet may also be a direct result of the increased plant cell and organic acid intake typical of herbivores (Houpt, 1989). Plant cells contain organic anions (citric, oxalic, malonic and fumaric acids) that are electrically balanced by potassium and other cations. Citrate and other organic anions are oxidised to  $\text{CO}_2$  and  $\text{H}_2\text{O}$ , but their breakdown requires hydrogen ions, which are derived from the hydration of carbon dioxide. Thus, as organic anions are oxidised,  $\text{HCO}_3^-$  is produced, which has an alkalotic effect on body fluid pH (Houpt, 1989). The possibility of counteracting the acidosis induced by intensive exercise by oral supplementation (generally with salts of  $\text{HCO}_3^-$ ) has been studied for a long time, but the effects on performance are not unequivocal (Kelso *et al.*, 1987; Lawrence *et al.*, 1987; Schuback *et al.*, 2002). This may be the first study to show that there is a natural way of achieving this effect, although a positive effect on performance remains to be proven.

Plasma glucose levels were higher during exercise in horses on the F diet. The origin and importance of this elevation in glucose concentration for exercise performance is not clear, but endurance during both maximal (Lacombe *et al.*, 2001) and sub-maximal (Farris *et al.*, 1998) exercise is improved by glucose infusion, indicating that high blood glucose availability could improve performance. Alterations in glucose metabolism (both in tissue uptake and release from the liver) are likely to occur in horses on forage-only diets, based on the low insulin levels observed here and earlier (Connysson *et al.*, 2010), and might be the reason for the elevated glucose levels during exercise. However, on the day after ETs, the plasma glucose concentration was lower in horses on the F diet and the NEFA concentration was higher. The importance of this is also unclear, but it might reflect the demand for glucose for resynthesis of glycogen and low dietary glucose availability (total daily intake of WSC and starch was  $\sim 860$  and  $3100$  g/day on diet F and FC, respectively) and increased utilisation of body fat for maintenance.

The low WSC intake might also have affected muscle glycogen synthesis (Lacombe *et al.*, 2004). Muscle glycogen content was lower ( $-13\%$ ) before and after ET in horses on the F diet, whereas relative glycogen depletion was similar. It is known that glycogen synthase is activated by insulin (Devlin and Horton, 1985) and the low insulin levels on the F diet might have been a limiting factor for glycogen synthesis. Further studies are needed to determine whether total muscle glycogen storage capacity is reduced on a forage-only diet or whether it is only the rate of synthesis that is reduced. It is important to note that all horses had trained  $\sim 96$  h before ET and that glycogen recovery might not have been complete. It is well documented that post-exercise recovery of glycogen stores takes several days in horses (Snow *et al.*, 1987; Hyypä *et al.*, 1997; Lacombe *et al.*, 2004). Although an exercise-induced reduction in muscle glycogen content of  $80\%$  has been shown to impair endurance during high-intensity exercise (Lacombe *et al.*, 2001), the importance of a smaller reduction, such as that induced by the F diet in the present study, is not known. Interestingly, oral acetate supplementation of a typical hay-grain diet has been shown to enhance the rate of glycogen resynthesis during the initial 4-hour recovery period after muscle glycogen depletion (Waller *et al.*, 2009), but our data suggest that increased acetate availability does not maximally support glycogen repletion between training sessions. However, a recent study (Essén-Gustavsson *et al.*, 2010) of horses on forage-only diets with different CP content showed that high CP forage increased muscle glycogen content compared with a forage providing the CP intake recommended by NRC (2007). This shows that CP is important for glycogen content and might have increased the glycogen content in horses on diet FC in the present study.

One argument against using forage-only or high-forage diets for performance horses is the risk of an unwanted increase in BW. This is probably based on the common perception that fibre is 'bulk' and perhaps also on knowledge of

the water-holding capacity of plant fibres. In a study on riding horses (Ellis *et al.*, 2002), a forage-only diet increased BW and heart rate during sub-maximal exercise, suggesting that performance might be impaired. However, it should be noted that the energy content of the forage in that study was not high enough to support the energy needs of horses in race training and, accordingly, the digestibility was not high enough, thereby causing 'bulk weight gain'. The present study showed no (ET days) or a limited (3 kg in pooled data) increase in BW in horses fed a high-energy forage diet. There was also no effect on heart rate and breathing frequency. It is likely that differences in chemical composition between forages are the reason for the differences in BW change in these studies. In the present study, the forage in diet F was early-cut and had high fibre digestibility, as reported by Ragnarsson and Jansson (2011) for the same batch of forage.

Part of the increase in BW observed on a forage-only diet could also be due to increased plasma volume, as indicated by the lowered TPP. It has been suggested that the hindgut serves as a fluid reservoir and that fibre-rich diets increase this reservoir (Meyer, 1987). The change in TPP with altered fibre intake could therefore reflect changes in the equilibrium between the gut and the extracellular fluid. It has been shown that increased forage intake lowers TPP (Danielsen *et al.*, 1995) and also that signs of dehydration (increased TPP) following feed deprivation are delayed in horses on a forage-only diet compared with those on a mixed diet (Connysson *et al.*, 2010). However, BW recovery was similar on both diets in the present study. The results also showed that it may take 2 to 3 days for horses transported to an exercise event to recover their BW. The loss of BW on ET day was due to lack of feed intake (less than  $60\%$  of the allowance) and to fluid and faeces losses during transportation and exercise.

There is also anecdotal information that forage intake capacity is limited in performance horses. Horses in the present study consumed forage corresponding to  $1.95\%$  of BW and four of the horses maintained similar body condition on both diets. It is possible that the other two horses would have been maintained on a forage diet with slightly higher energy content (around  $11$  MJ metabolisable energy/kg DM). In all horses on the forage-only diet, the maintenance of energy intake and body condition was to some extent due to selection of the forage offered, as reflected in lower digestibility of leftovers from this diet. This contributed to higher energy intake on the forage-only diet and was probably an attempt to maximise energy intake.

Another concern about a forage-dominated diet for athletic horses is the increased heat increment of feeding. In the present study, this was probably reflected in the higher water intake on diet F. However, body temperature was similar, as would have been expected in a homeothermic animal, and it was not possible to draw any conclusions about evaporative fluid losses during exercise from the results, as only BW was measured before and after exercise (including sweat, faecal and respiratory losses).

No feed-related clinical or behavioural disturbances very observed during the study. Exercise temperament, novel

object reaction, voluntary motion and post-exercise feeding (Jansson, 2010) were also evaluated in the present study, and numerical, but not statistically significant, differences were detected, with the exception of observations on post-exercise feed intake, which was more common on diet F. However, subjectively more aggression was observed in horses fed diet FC, especially in the afternoon when horses were fetched from the paddock (one at a time) to be put in their boxes, where feed was available. Therefore, our impression is that the FC diet might have affected behaviour (making horses more active and reactive), although the recording system used could not verify this.

In conclusion, the present study indicates that a high-energy, forage-only diet alters the metabolic response to exercise and, with the exception of lowered glycogen stores, appears to have positive rather than negative effects on performance traits of Standardbred horses.

### Acknowledgements

The authors thank HästKraft AB and Lantmännen Box for providing the forages, and trainer Anna Svensson and the staff at Mälarkliniken, Sigtuna, Sweden, for assistance during the study. This research received financial support from Stiftelsen Svensk Hästforskning, Sweden.

### References

Connysson M, Essén-Gustavsson B, Lindberg JE and Jansson A 2010. Effects of feed deprivation on Standardbred horses in training fed a forage-only diet and a 50:50 forage-oats diet. *Equine Veterinary Journal Supplement* 38, 335–340.

Connysson M, Muhonen S, Lindberg JE, Essén-Gustavsson B, Nyman G, Nostell K and Jansson A 2006. Effects on exercise response, fluid and acid-base balance of protein intake from forage-only diets in Standardbred horses. *Equine Veterinary Journal Supplement* 36, 648–653.

Danielsen K, Lawrence LM, Siciliano P, Powell D and Thomson K 1995. Effects of diet on weight and plasma variables in endurance exercised horses. *Equine Veterinary Journal Supplement* 18, 372–377.

Devlin JT and Horton ES 1985. Effects of prior high-intensity exercise on glucose metabolism in normal and insulin-resistant men. *Diabetes* 34, 973–979.

Ellis JM, Hollands T and Allen DE 2002. Effect of forage on body weight and performance. *Equine Veterinary Journal Supplement* 34, 66–70.

Essén-Gustavsson B, Connysson M and Jansson A 2010. Effects of crude protein intake from forage-only diets on muscle amino acids and glycogen levels in horses in training. *Equine Veterinary Journal* 38, 341–346.

Essén-Gustavsson B, McMiken D, Karlstrom K, Lindholm A, Persson S and Thornton J 1989. Muscular adaptation of horses during intensive training and detraining. *Equine Veterinary Journal* 21, 27–33.

Farris JW, Hinchcliff KW, McKeever KH, Lamb DR and Thomson DL 1998. Effects of tryptophan and of glucose on exercise capacity of horses. *Journal of Applied Physiology* 85, 807–816.

Geor RJ 2006. The role of nutritional supplements and feeding strategies in equine athletic performance. *Equine and Comparative Exercise Physiology* 3, 109–119.

Gillham SB, Dodman NH, Shuster L, Kream RK and Rand W 1994. The effect of diet on cribbing behaviour and plasma  $\beta$ -endorphins in horses. *Applied Animal Behaviour Science* 41, 147–153.

Glade MJ 1983. Nutrition and performance of racing Thoroughbreds. *Equine Veterinary Journal* 15, 31–36.

Harris PA and Harris RC 2005. Ergogenic potential of nutritional strategies and substances in horses. *Livestock Production Science* 92, 147–165.

Hintz HF, Argenzio RA and Schryver HF 1971. Digestion coefficients, blood glucose levels and molar percentages of VFA in intestinal fluid of ponies fed varying forage-grain ratios. *Journal of Animal Science* 33, 992–995.

Houpt RT 1989. Water balance and excretion. In *Dukes's physiology of domestic animals*, 10th edition (ed. MJ Swenson), pp. 496. Comstock, Cornell University Press, Ithaca, New York, USA.

Hudson JM, Cohen ND, Gibbs PG and Thompson JA 2001. Feeding practices associated with colic in horses. *Journal of the American Veterinary Medical Association* 219, 1419–1425.

Hyypää S, Räsänen LA and Pösö AR 1997. Resynthesis of glycogen in skeletal muscle from Standardbred trotters after repeated bouts of exercise. *American Journal of Veterinary Research* 58, 162–166.

Jansson A 2010. Effects of diet on behaviour of Standardbred horses in training. In *The impact of nutrition on the health and welfare of horses* (ed. AD Ellis, AC Longland, M Coenen and N Miraglia), Part 2, 88p, Wageningen Academic Publishers, The Netherlands.

Kelso TB, Hodgson DR, Witt EH, Bayly WM, Grant BD and Gollnick PD 1987. Bicarbonate administration and muscle metabolism during high-intensity exercise. In *Equine exercise physiology 2* (ed. JR Gillespie and NE Robinson), pp. 438–447. ICEEP Publications, Davis, CA, USA.

Lacombe VA, Hinchcliff KW, Geor RJ and Baskin CR 2001. Muscle glycogen depletion and subsequent replenishment affect anaerobic capacity of horses. *Journal of Applied Physiology* 91, 1782–1790.

Lacombe VA, Hinchcliff KW, Kohn CW, Devor ST and Taylor LE 2004. Effects of feeding meals with various soluble-carbohydrate content on muscle glycogen synthesis after exercise in horses. *American Journal of Veterinary Research* 65, 916–923.

Lawrence LM, Miller PA, Bechtel PJ, Kane RA, Kurcz EV and Smith JS 1987. The effect of sodium bicarbonate ingestion on blood parameters in exercising horses. In *Equine exercise physiology 2* (ed. JR Gillespie and NE Robinson), pp. 448–455. ICEEP Publications, Davis, CA, USA.

Lindgren E 1979. The nutritional value of roughages determined in vivo and by laboratory methods. Report 45: 63, Department of Animal Nutrition and Management, Swedish University of Agricultural Sciences, Sweden.

Lindholm A and Piehl K 1974. Fibre composition, enzyme activity and concentrations of metabolites and electrolytes in muscles of standardbred horses. *Acta Veterinaria Scandinavica* 15, 287–309.

Lowry OH and Passonneau JV 1973. A flexible system for enzymatic analysis, pp. 1–291. Academic Press, NY, USA.

Luthersson N, Nielsen KH, Harris P and Parkin TD 2009. Risk factors associated with equine gastric ulceration syndrome (EGUS) in 201 horses in Denmark. *Equine Veterinary Journal* 41, 625–630.

MacLeay JM, Sorum SA, Marsh WE and Sorum MD 1999. Epidemiologic analysis of factors influencing exertional rhabdomyolysis in Thoroughbreds. *American Journal of Veterinary Research* 60, 1562–1566.

Meyer H 1987. Nutrition of the equine athlete. In *Equine exercise physiology 2* (ed. JR Gillespie and NE Robinson), pp. 644–673. ICEEP Publications, Davis, CA, USA.

Muhonen S, Lindberg J E, Bertilsson J and Jansson A 2009. Effects on fluid balance and exercise response in Standardbred horses feed silage, haylage and hay. *Comparative Exercise Physiology* 5, 133–142.

National Research Council (NRC) 1989. Nutrient requirements of horses, 5th edition. National Academic Press, Washington, DC, USA.

NRC 2007. Nutrient Requirements of Horses, 6th edition. National Academic Press, Washington, DC, USA.

Pagan J, Geor RJ, Harris PA, Hoekstra K, Gardner S, Hudson C and Prince A 2002. Effects of fat adaptation on glucose kinetics and substrate oxidation during low-intensity exercise. *Equine Veterinary Journal Supplement* 34, 33–38.

Palmgren-Karlsson C, Lindberg JE and Rundgren M 2000. Associative effects on total tract digestibility in horses fed different ratios of grass hay and whole oats. *Livestock Production Science* 65, 143–153.

Palmgren-Karlsson C, Jansson A, Essén-Gustavsson B and Lindberg JE 2002. Effect of molassed sugar beet pulp on nutrient utilisation and metabolic parameters during exercise. *Equine Veterinary Journal Supplement* 34, 44–49.

Persson SGB 1983. Analysis of fitness and state of training: evaluation of exercise tolerance and fitness in the performance horse. In *Equine exercise physiology 1* (ed. DH Snow, SGB Persson and RJ Rose), pp. 441–457. Granta Publications Cambridge, UK.

- Potter GD, Hughes SL, Julen TR and Swinney DDL 1992. A review of research on digestion and utilization of fat by the equine. *Pferdeheilkunde Sonderausgabe* 119–123.
- Ragnarsson S and Jansson A 2011. A comparison of grass haylage digestibility and metabolic plasma profile in Icelandic and Standardbred horses. *Journal of Animal Physiology and Animal Nutrition* 95, 273–279.
- Redbo I, Redbo-Torstensson P, Ödberg FO, Hedendahl A and Holm J 1998. Factors affecting behavioural disturbances in race-horses. *Journal of Animal Science* 66, 475–481.
- Schuback K, Essén-Gustavsson B and Persson SG 2002. Effect of sodium bicarbonate administration on metabolic responses to maximal exercise. *Equine Veterinary Journal Supplement* 34, 539–544.
- Snow DH, Harris RC, Harman JC and Marlin DJ 1987. Glycogen repletion following different diets. In *Equine exercise physiology 2* (ed. JR Gillespie and NE Robinson), pp. 701–710. ICEEP Publications, Davis CA, USA.
- Tinker MK, White NA, Lessard P, Thatcher CD, Pelzer KD, Davis B and Carme DK 1997. Prospective study of equine colic risk factors. *Equine Veterinary Journal* 29, 454–458.
- Voiton DM, Navet R, Lacombe VA, Sluse F, Essen-Gustavsson B, Hinchcliff KW, Rivero JLL, Serteyn D and Valberg S 2007. Muscle energetic in exercising horses. *Equine and Comparative Exercise Physiology* 4, 105–118.
- Waller AP, Geor RJ, Spriet LL, Heighenhasuer GJ and Lindinger MI 2009. Oral acetate supplementation after prolonged moderate intensity exercise enhances early muscle glycogen resynthesis in horses. *Experimental Physiology* 94, 888–898.
- Waller A and Lindinger M 2007. The effect of oral sodium acetate administration on plasma acetate concentration and acid–base state in horses. *Acta Veterinaria Scandinavica* 49, 38, doi:10.1186/1751-0147-49-38.
- Waters AJ, Nicol CJ and French NP 2002. Factors influencing the development of stereotypic and redirected behaviours in young horses: findings of a four year prospective epidemiological study. *Equine Veterinary Journal* 34, 572–579.
- Willard JG, Willard JC, Wolfram SA and Baker JP 1977. Effect of diet on cecal pH and feeding behavior of horses. *Journal of Animal Science* 45, 87–93.
- Williamson A, Rogers CW and Firth EC 2007. A survey of feeding, management and faecal pH of Thoroughbred racehorses in the North Island of New Zealand. *New Zealand Veterinary Journal* 55, 337–341.
- Willing B, Vörös A, Roos S, Jones C, Jansson A and Lindberg JE 2009. Changes in faecal bacteria associated with concentrate and forage-only diets fed to horses in training. *Equine Veterinary Journal* 41, 908–914.
- Wilson RG, Thornton JR, Inglis S and Ainscow J 1987. Skeletal muscle adaptation in racehorses following high intensity interval training. In *Equine exercise physiology 2* (ed. JR Gillespie and NE Robinson), pp. 367–375. ICEEP Publications, Davis CA, USA.



Reproduced with permission of the copyright owner. Further reproduction prohibited without permission.